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| (54) Title: INTESTINAL TREFOIL PROTEINS (57) Abstract <p>Intestinal trefoil factors and nucleic acids encoding intestinal trefoil factors are disclosed. The intestinal trefoil factors disclosed are resistant to destruction in the digestive tract and can be used for the treatment of peptic ulcer diseases, inflammatory bowel diseases and other insults.</p> | | |

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- 1 -

INTESTINAL TREFOIL PROTEINSBackground

The field of the invention is peptides that are
5 useful for the diagnosis, prevention, or treatment of
wounds, including those that are associated with a
gastrointestinal disorder.

Jørgensen et al. (*Regulatory Peptides* 3:231, 1982)
describe a porcine pancreatic peptide, pancreatic
10 spasmolytic peptide (PSP). PSP was found to inhibit
"gastrointestinal motility and gastric acid secretion in
laboratory animal after parenteral as well as oral
administration." It was suggested that "if the results
in animal experiments can be confirmed in man, PSP may
15 possess a potential utility in treatment of
gastroduodenal ulcer diseases."

Summary of the Invention

In a first aspect, the invention features a
purified nucleic acid encoding an intestinal trefoil
20 factor (ITF).

In preferred embodiments, the intestinal trefoil
factor is mammalian intestinal trefoil factor, preferably
human, rat, bovine, or porcine intestinal trefoil factor.
In another preferred embodiment, the purified nucleic
25 acid encoding an intestinal trefoil factor is present
within a vector.

In a related aspect, the invention features a cell
that includes a vector encoding an intestinal trefoil
factor.

30 In another related aspect, the invention features
a substantially pure intestinal trefoil factor. In a
preferred embodiment, the polypeptide is detectably
labelled. In a related aspect, the invention features a
therapeutic composition that includes an intestinal

- 2 -

trefoil factor and a pharmacologically acceptable carrier.

In another aspect, the invention features a monoclonal antibody which preferentially binds (i.e.,
5 forms an immune complex with) an intestinal trefoil factor. In a preferred embodiment, the monoclonal antibody is detectably labelled.

In a related aspect, the invention features a method for detecting human intestinal trefoil factor in a
10 human patient. The method includes the steps of contacting a biological sample obtained from the patient with a monoclonal antibody which preferentially binds intestinal trefoil factor, and detecting immune complexes formed with the monoclonal antibody. In preferred
15 embodiments the biological sample is an intestinal mucosal scraping, or serum.

In a related aspect, the invention features a method for treating digestive disorders in a human patient, which method involves administering to the
20 patient a therapeutic composition that includes an intestinal trefoil factor and a pharmacologically acceptable carrier. Additional disorders that can also be treated are described below.

In another aspect, the invention features a method
25 for detecting binding sites for intestinal trefoil factor in a patient. The method involves contacting a biological sample obtained from the patient with the factor, and detecting the factor bound to the biological sample as an indication of the presence of the binding
30 sites in the sample. By "binding sites," as used herein, is meant any antibody or receptor that binds to an intestinal trefoil factor protein, factor, or analog. The detection or quantitation of binding sites may be a useful reflection of abnormalities of the
35 gastrointestinal tract.

- 3 -

In another aspect, the invention features substantially pure trefoil factor. In preferred embodiments, the intestinal trefoil factor is human, porcine, or bovine trefoil factor.

5 By "intestinal trefoil factor" ("ITF") is meant any protein that is substantially homologous to rat intestinal trefoil factor (Fig. 2; SEQ ID NO:2) and which is expressed in the large intestine, small intestine, or colon to a greater extent than it is expressed in tissues
10 other than the small intestine, large intestine, or colon. Also included are: allelic variations; natural mutants; induced mutants; proteins encoded by DNA that hybridizes under high or low stringency conditions to ITF encoding nucleic acids retrieved from naturally occurring
15 material; and polypeptides or proteins retrieved by antisera to ITF, especially by antisera to the active site or binding domain of ITF. The term also includes other chimeric polypeptides that include an ITF.

The term ITF also includes analogs of naturally
20 occurring ITF polypeptides. Analogs can differ from naturally occurring ITF by amino acid sequence differences or by modifications that do not affect sequence, or by both. Analogs of the invention will generally exhibit at least 70%, more preferably 80%, more
25 preferably 90%, and most preferably 95% or even 99%, homology with all or part of a naturally occurring ITF sequence. The length of comparison sequences will generally be at least 8 amino acid residues, usually at least 20 amino acid residues, more usually at least 24
30 amino acid residues, typically at least 28 amino acid residues, and preferably more than 35 amino acid residues. Modifications include *in vivo*, or *in vitro* chemical derivatization of polypeptides, e.g., acetylation, or carboxylation. Also included are
35 modifications of glycosylation, e.g., those made by

- 4 -

modifying the glycosylation patterns of a polypeptide during its synthesis and processing or in further processing steps, e.g., by exposing the polypeptide to enzymes that affect glycosylation derived from cells that normally provide such processing, e.g., mammalian glycosylation enzymes. Also embraced are versions of the same primary amino acid sequence that have phosphorylated amino acid residues, e.g., phosphotyrosine, phosphoserine, or phosphothreonine. Analogs can differ from naturally occurring ITF by alterations of their primary sequence. These include genetic variants, both natural and induced. Induced mutants may be derived by various techniques, including random mutagenesis of the encoding nucleic acids using irradiation or exposure to ethanemethylsulfate (EMS), or may incorporate changes produced by site-specific mutagenesis or other techniques of molecular biology. See, Sambrook, Fritsch and Maniatis (1989), Molecular Cloning: A Laboratory Manual (2d ed.), CSH Press, Cold Spring Harbor, New York. Also included are analogs that include residues other than naturally occurring L-amino acids, e.g., D-amino acids or non-naturally occurring or synthetic amino acids, e.g., β or γ amino acids.

In addition to substantially full-length polypeptides, the term ITF, as used herein, includes biologically active fragments of the polypeptides. As used herein, the term "fragment," as applied to a polypeptide, will ordinarily be at least 10 contiguous amino acids, typically at least 20 contiguous amino acids, more typically at least 30 contiguous amino acids, usually at least 40 contiguous amino acids, preferably at least 50 contiguous amino acids, and most preferably at least 60 to 80 or more contiguous amino acids in length. Fragments of ITF can be generated by methods known to those skilled in the art. The ability of a candidate

- 5 -

fragment to exhibit a biological activity of ITF can be assessed by methods known to those skilled in the art.

Also included in the term "fragment" are biologically active ITF polypeptides containing amino acids that are normally removed during protein processing, including additional amino acids that are not required for the biological activity of the polypeptide, or including additional amino acids that result from alternative mRNA splicing or alternative protein processing events.

An ITF polypeptide, fragment, or analog is biologically active if it exhibits a biological activity of a naturally occurring ITF, e.g., the ability to alter gastrointestinal motility in a mammal.

The invention also includes nucleic acid sequences, and purified preparations thereof, that encode the ITF polypeptides described herein, as well as antibodies, preferably monoclonal antibodies, that bind specifically to ITF polypeptides.

As used herein, the term "substantially pure" describes a compound, e.g., a nucleic acid, a protein, or a polypeptide, e.g., an ITF protein or polypeptide, that is substantially free from the components that naturally accompany it. Typically, a compound is substantially pure when at least 60%, more preferably at least 75%, more preferably at least 90%, and most preferably at least 99%, of the total material (by volume, by wet or dry weight, or by mole per cent or mole fraction) in a sample is the compound of interest. Purity can be measured by any appropriate method, e.g., in the case of polypeptides, by column chromatography, polyacrylamide gel electrophoresis, or HPLC analysis.

By "isolated DNA" is meant DNA that is free of the genes which, in the naturally-occurring genome of the organism from which the given DNA of the invention is

- 6 -

derived, flank the DNA. The term "isolated DNA" thus encompasses, for example, cDNA, cloned genomic DNA, and synthetic DNA. A "purified nucleic acid," as used herein, refers to a nucleic acid sequence that is substantially free of other macromolecules (e.g., other nucleic acids and proteins) with which it naturally occurs within a cell. In preferred embodiments, less than 40% (and more preferably less than 25%) of the purified nucleic acid preparation consists of such other macromolecule.

"Homologous," as used herein, refers to the subunit sequence similarity between two polymeric molecules, e.g., between two nucleic acid molecules, e.g., two DNA molecules, or two polypeptide molecules. When a subunit position in both of the molecules is occupied by the same monomeric subunit, e.g., if a position in each of two DNA molecules is occupied by adenine, then they are homologous at that position. The homology between two sequences is a direct function of the number of matching or homologous positions, e.g., if half, e.g., 5 of 10, of the positions in two compound sequences are homologous then the two sequences are 50% homologous; if 90% of the positions, e.g., 9 of 10, are matched or homologous the two sequences share 90% homology. By way of example, the DNA sequences 5'-ATTGCC-3' and 5'-TATGGC-3' share 50% homology. By "substantially homologous" is meant largely but not wholly homologous.

The ITF proteins of the invention are resistant to destruction in the digestive tract, and can be used for treatment of peptic ulcer diseases, inflammatory bowel diseases, and for protection of the intestinal tract from injury caused by insults such as radiation injury or bacterial infection. An ITF protein, fragment, or analog can also be used to treat neoplastic cancer and, as

- 7 -

described further below, to protect (i.e., by inhibiting the formation of lesions) or to treat any part of the body from inflammation or injuries such as lesions, ulcerations, burns, or abrasions.

5 In general, trefoil proteins, including ITF, are useful for the treatment of disorders of and damage to the alimentary canal, including the mouth, esophagus, stomach, and large and small intestine, as well as for the protection and treatment of tissues that lie outside
10 the alimentary canal. The polypeptide can be used either to treat lesions in these areas or to inhibit the formation of lesions. The latter tissues include, for example, the external surface of the skin, the surface of the eye, the mucosa of the nasal passages and respiratory
15 tract, and the genitourinary tract.

One of the most common bacterial infections is caused by *Helicobacter pylori* (*H. pylori*), which leads to active, chronic gastritis and frequently to associated syndromes such as duodenal ulcer, gastric ulcer, gastric
20 cancer, MALT lymphoma, or Menetrier's syndrome. Eradication of *H. pylori* has been shown to reduce the recurrence of duodenal and gastric ulcers. Furthermore, it has been postulated that widespread treatment of *H. pylori* will reduce the incidence of gastric carcinoma,
25 which is the second leading cause of cancer related death world-wide.

Long-standing gastritis associated with *H. pylori* infection is often associated with the expression of intestinal-like features in the gastric mucosa. This
30 condition, referred to as intestinal metaplasia (IM), may signal an increased risk of gastric cancer. The etiology of IM is unclear; it could represent a mutational adaptation or defense against *H. pylori* infection. For example, the metaplastic mucosa may produce mucus or
35 other substances that create an environment that is

- 8 -

hostile to *H. pylori*. ITF can be used in the treatment of *H. pylori* infection and conditions associated with *H. pylori* infection (e.g., ulcers, gastric carcinoma, non-ulcer dyspepsia, gastritis, and esophageal lesions associated with gastro-esophageal reflux disease). ITF is useful for treatment of these conditions because of its generally protective effect on the gastrointestinal tract. In addition, ITF promotes the maintenance of mucosal integrity. ITF can be used to inhibit adhesion to or colonization of the mucosa by *H. pylori*. In this application, ITF or fragments or variants thereof which inhibit adhesion or colonization of the mucosa by *H. pylori* are useful. Such molecules can be identified using assays known to those skilled in the art, including the *H. pylori* binding assay described below.

ITF may also be used promote healing of tissues damaged by conditions associated with *H. pylori* infection. In this regard, it is important that addition of trefoil proteins to wounded monolayers of confluent intestinal epithelial cells increases the rate of epithelial cell migration into the wound. This effect is enhanced by concomitant addition of mucin glycoproteins, the other dominant product of goblet cells.

Just as ITF can be used to protect other parts of the gastro-intestinal tract or alimentary canal, such as the intestine, it can be used to protect the mouth and esophagus from damage caused by radiation therapy or chemotherapy. ITF can also be used to protect against (i.e., inhibit the injury caused by) and/or to treat damage caused by alcohols or drugs generally. Additional tissues that can be protected or treated by an ITF include those listed above, which lie outside the alimentary canal.

Members of the trefoil family, including ITF, can be used in the treatments discussed above. Skilled

- 9 -

artisans may review these proteins in Sands et al. (1996, *Ann. Rev. Physiol.* 58:253-273). As stated above, the invention encompasses biologically active fragments of the trefoil proteins. Fragments that retain the trefoil structure (i.e., the three loop structure) or that lie within regions of the protein that are highly conserved may prove particularly useful. Thus, such fragments can encompass portions of ITF from about the first cysteine residue involved in a disulfide bond of the three loop structure to about the last cysteine residue involved in a disulfide bond of the three loop structure.

Variants of a selected trefoil protein are least 60%, preferably at least 75%, more preferably at least 90%, and most preferably at least 95% identical to the selected trefoil protein, preferably a human trefoil protein, more preferably human ITF.

The term "identical," as used herein in reference to polypeptide or DNA sequences, refers to the subunit sequence identity between two molecules. In the case of amino acid sequences that are less than 100% identical to a reference sequence, the non-identical positions are preferably, but not necessarily, conservative substitutions for the reference sequence. Conservative substitutions typically include substitutions within the following groups: glycine, alanine; valine, isoleucine, leucine; aspartic acid, glutamic acid; asparagine, glutamine; serine, threonine; lysine, arginine; and phenylalanine, tyrosine. Sequence identity is typically measured using sequence analysis software such as the Sequence Analysis Software Package of the Genetics Computer Group at the University of Wisconsin (Biotechnology Center, 1710 University Avenue, Madison, WI 53705), and the default parameters specified therein.

A variant of a selected trefoil protein preferably has the amino acids present in the naturally-occurring

- 10 -

form of the selected trefoil protein at the more highly conserved amino acid positions of the protein. Thus, a variant of human ITF preferably is identical to naturally-occurring human ITF at all or nearly all of the more highly conserved positions. Sequence conservation among trefoil proteins is evident in Table 1 of Sands et al. (*supra*) which can be used by those skilled in the art to identify more conserved residues.

The invention features a method for treating or inhibiting the formation of lesions in the alimentary canal of a patient by administering to the patient at least one trefoil polypeptide, or a biologically active fragment thereof. The lesions typically occur in the mucosa of the alimentary canal, and may be present in the mouth, esophagus, stomach, or intestine of the patient. The lesions can be caused in several ways. For example, the patient may be receiving radiation therapy or chemotherapy for the treatment of cancer. These treatments typically cause lesions in the mouth and esophagus of the patient. Skilled artisans will recognize that it may be useful to administer the proteins of the invention to the patient before such treatment is begun. Alternatively, the lesions can be caused by: (1) any other drug, including alcohol, that damages the alimentary canal, (2) accidental exposure to radiation or to a caustic substance, (3) an infection, or (4) a digestive disorder including but not limited to non-ulcer dyspepsia, gastritis, peptic or duodenal ulcer, gastric cancer, MALT lymphoma, Menetrier's syndrome, gastro-esophageal reflux disease, and Crohn's disease.

Tissues that lie outside the alimentary canal can also be treated by administering to the patient at least one trefoil peptide, or a biologically active fragment thereof, in the event those tissues are damaged by inflammation, a lesion, ulcer, abrasion, burn, or other

- 11 -

wound, or are at risk of being so injured (i.e., the method can be carried out prophylactically).

The peptide that is administered may be any peptide in the trefoil family, such as intestinal trefoil peptide (ITF), spasmolytic peptide (SP), and pS2. For the treatment of human patients it is expected that the peptide will be expressed by a human gene. However, eucaryotic trefoil peptides, such as those cloned from the rat and mouse genomes may also prove effective.

10 These peptides may be isolated from a naturally occurring source or synthesized by recombinant techniques. It is expected that the typical route of administration will be oral. Determining other routes of administration, and the effective dosage are well within the skills of

15 ordinary artisans and will depend on many factors known to these artisans. The trefoil proteins may be administered singly, in combination with one another, and/or in combination with mucin glycoprotein preparations. "Treatment of lesions" encompasses both

20 the inhibition of the formation of lesion and the healing of lesions already formed. Biologically active fragments and variants of a trefoil protein, particularly ITF, which promote healing of lesions or inhibit the formation of lesions, are useful in the treatments of the

25 invention.

The polypeptides of the invention can also be used for diagnostic purposes. For example, the polypeptides can be used in an assay to quantitate intestinal trefoil factor and related polypeptides, such as fragments and

30 analogs, in tissues, serum, and other biological samples. In many inflammatory bowel diseases, and potentially many other inflammatory conditions, the expression of ITF is reduced. Thus, areas where expression is relatively low indicate the presence of injured tissue.

- 12 -

Alternatively, the polypeptides can be bound to a diagnostic marker and administered to a patient. In this circumstance, the polypeptide would facilitate the distribution of the diagnostic marker within any tissue that expresses a receptor for the polypeptide. The diagnostic marker can be any substance that is capable of being detected. Those of skill in the art are well aware of numerous imaging agents that can be used according to the present invention.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof, and from the claims.

Brief Description of the Drawings

Figure 1 is a depiction of the nucleotide sequence of rat trefoil factor (SEQ ID NO:1).

Figure 2 is a depiction of the deduced amino acid sequence of rat trefoil factor (SEQ ID NO:2).

Figure 3 is a depiction of the amino acid sequences of rat trefoil factor, pS2 protein, and pancreatic spasmodolytic polypeptide (SP). The sequences are aligned to illustrate the amino acid sequence homology between the proteins. Dashes (-) indicate the insertion of spaces which optimize alignment. Bars indicate sequence identity.

Figure 4 depicts the disulfide bond structure proposed for pS2 (SEQ ID NO:15; panel A) and PSP (SEQ ID NO:16; panel B).

Figure 5 is a depiction of the proposed disulfide bond structure of rat intestinal trefoil factor (SEQ ID NO:17).

Figure 6 is a depiction of the nucleotide sequence of the human intestinal trefoil factor cDNA and the corresponding deduced amino acid sequence (SEQ ID NO:3).

- 13 -

Figure 7 is a diagram depicting the strategy used to mutate the ITF gene in embryonic stem cells.

Figure 8 is a graph depicting survival following administration of Dextran Sulfate Sodium (DSS; 2.5% w/v in drinking water for 9 consecutive days), shown as Kaplan-Meier transform of probability versus days of DSS treatment.

Detailed Description

Purification and cloning of rITF

10 An inhibitor of soft agar colony formation by human breast carcinoma-derived BT-20 cells (ATTC HTB79) was isolated from cytology-positive human malignant effusions (Podolsky et al., *Cancer Res.* 48:418, 1988). The factor also inhibited soft agar colony formation by
15 human colon carcinoma-derived HCT15 cells (ATTC-CCL225). Inhibition was not observed for polyoma and murine sarcoma virus transformed rodent fibroblast lines. The isolated factor (transformed cell-growth inhibiting factor or TGIF) had an apparent molecular weight of
20 110,000 Da and appeared to consist of two 55,000 Da subunits linked by sulfhydryl bonds.

The purified protein was partially sequenced. The sequence from the amino terminal 14 amino acids was used to produce a set of degenerate oligonucleotide probes for
25 screening of a rat intestinal epithelial cell cDNA library.

A rat intestinal cDNA library (Lambda ZAP[®] II, Stratagene, La Jolla, CA) was produced by standard techniques (Ausubel et al., Eds., *In Current Protocols in*
30 *Molecular Biology*, John Wiley & Sons, New York, 1989) using cells purified by the method of Weisner (*J. Biol. Chem.* 248:2536, 1973). Screening of the cDNA library with the fully degenerate oligonucleotide probe described above resulted in the selection of 21 clones. One of the

- 14 -

clones (T3411) included a core sequence which encoded a single open reading frame. The nucleotide sequence of the open reading frame and flanking DNA is presented in Fig. 1 (SEQ ID NO:1). The insert present in T3411 was nick translated (Ausubel et al., *supra*) to produce a radioactively labelled probe for Northern blot analysis of rat poly(A)⁺ RNA. Northern analysis demonstrated that RNA corresponding to the cloned cDNA fragment was expressed in small intestine, large intestine, and kidney; no expression was detected in the lung, spleen, heart, testes, muscle, stomach, pancreas, or liver. In the tissues in which the RNA was expressed, the level was comparable to that of actin.

The open reading frame of clone T3411 encoded an 81 amino acid peptide (Fig. 2; SEQ ID NO:2). Comparison of the sequence of the encoded peptide, referred to as rat intestinal trefoil factor (rITF), to the sequence of proteins in the Genbank database revealed significant homology to human breast cancer associated peptide (pS2; Jakowlev et al., *Nucleic Acids Res.* 12:2861, 1984) and porcine pancreatic spasmolytic peptide (PSP; Thim et al., *Biochem. Biophys. Acta.* 827:410, 1985). Figure 3 illustrates the homology between rITF, PSP, and pS2. Porcine pancreatic spasmolytic factor (PSP) and pS2 are both thought to fold into a characteristic structure referred to as a trefoil. A trefoil structure consists of three loops formed by three disulfide bonds. pS2 is thought to include one trefoil (Fig. 4A), and PSP is thought to include two trefoils (Fig. 4B). The region of rITF (nucleotide 114 to nucleotide 230 which encodes cysto phe), which is most similar to PSP and pS2, includes six cysteines, all of which are in the same position as the cysteines which make up the trefoil in pS2 (Fig. 3). Five of these six cysteines are in the same position as the cysteines which form the amino terminal trefoil of

- 15 -

PSP (Fig. 3). Figure 5 depicts the proposed disulfide bond configuration of rITF.

Based on homology to PSP and ps2 (Mori et al., *Biochem. Biophys. Res. Comm.* 155:366, 1988; Jakowlew et al., *Nucleic Acids Res.* 12:2861, 1984), rITF includes a presumptive pro-sequence (met1 to ala22) in which 12 of 22 amino acids have hydrophobic side chains.

Production of Anti-rITF Antibodies

A peptide corresponding to the carboxy-terminal 21 amino acids of rITF was synthesized and coupled to bovine serum albumin (BSA). This conjugate (and the unconjugated peptide) was used to raise polyclonal antibodies in rabbits. All procedures were standard protocols such as those described in Ausubel et al. (supra). The anti-rITF antibodies were used in an indirect immunofluorescence assay for visualization of rITF in rat tissues. Cryosections of rat tissues were prepared using standard techniques, and fluorescein labelled goat anti-rabbit monoclonal antibody (labelled antibodies are available from such suppliers Kirkegaard and Perry Laboratories, Gaithersburg, MD; and Bioproducts for Science, Inc., Indianapolis, IN) was used to detect binding of rabbit anti-rITF antibodies. By this analysis rITF appears to be present in the goblet cells of the small intestine but not in the stomach or the pancreas.

Cloning of Human Intestinal Trefoil Factor

DNA encoding the rat intestinal trefoil factor can be used to identify a cDNA clone encoding the human intestinal trefoil factor (hITF). This can be accomplished by screening a human colon cDNA library with a probe derived from rITF or with a probe derived from part of the hITF gene. The latter probe can be obtained from a human colon or intestinal cDNA using the

- 16 -

polymerase chain reaction to isolate a part of the hITF gene. This probe can then serve as a specific probe for the identification of clones encoding all of the hITF gene.

5 Construction of a cDNA Library

A human colon or intestinal cDNA library in λ gt10 or λ gt11, or some other suitable vector is useful for isolation of hITF. Such libraries may be purchased (Clontech Laboratories, Palo Alto, CA: HLI034a, 10 HLI0346b). Alternatively, a library can be produced using mucosal scrapings from human colon or intestine. Briefly, total RNA is isolated from the tissue essentially as described by Chirgwin et al. (*Biochemistry* 18:5294, 1979; see also Ausubel et al., *supra*). An oligo 15 (dT) column is then used to isolate poly(A)⁺ RNA by the method of Aviv et al. (*J. Mol. Biol.* 134:743, 1972; see also Ausubel et al., *supra*). Double-stranded cDNA is then produced by reverse transcription using oligo (dT)₁₂₋₁₈ or random hexamer primers (or both). RNaseH and *E. coli* DNA 20 polI are then used to replace the RNA strand with a second DNA strand. In a subsequent step, *E. coli* DNA ligase and T4 DNA polymerase are used to close gaps in the second DNA strand and create blunt ends. Generally, the cDNA created is next methylated with *EcoRI* methylase 25 and *EcoRI* linkers are added (other linkers can be used depending on the vector to be used). In subsequent steps the excess linkers are removed by restriction digestion and the cDNA fragments are inserted into the desired vector. See Ausubel et al., *supra* and Sambrook et al. (*In* 30 *Molecular Cloning: A Laboratory Manual*, CSH Laboratory Press, Cold Spring Harbor, NY, 1990) for detailed protocols.

- 17 -

Useful vectors include: λ gt11, λ gt10, Lambda ZAP® II vector, Lambda Uni-ZAP™ XR vector, all available from Stratagene (La Jolla, CA).

The cDNA library must be packaged into phage; this is most readily accomplished by use of a commercial in vitro packaging kit, e.g., Gigapack® II Gold or Gigapack® II Plus (Stratagene, La Jolla, CA). See Ausubel et al. (supra) for packaging protocols and suitable host strains. The library is preferably amplified soon after packaging; this step generates sufficient clones for multiple screening of the library. See Ausubel et al. supra or Sambrook et al. supra for details of amplification protocols and procedures for storing the amplified library.

15 Screening of the cDNA Library

To screen the library it must be placed on an appropriate host strain (e.g., Y1090 or Y1088 for λ gt10 libraries, C600hflA for λ gt10 libraries). After plating the phage, plaques are transferred to nitrocellulose or nylon filters (See Ausubel et al., supra and Sambrook et al. supra). The filters are then probed with $\alpha^{32}\text{P}$ -labelled nick translated probe derived from rITF. The probe is preferentially generated using a portion of the region of rITF DNA coding for the trefoil structure (nucleotides 25 114 to 230 of SEQ ID NO:1, which encode cys32 to phe71 of SEQ ID NO:2). This region is conserved between rITF, pS2 and PSP, and it is likely that this region is conserved between rITF and hITF. Once a plaque is identified, several cycles of plaque purification are required to isolate a pure clone encoding hITF. A phage DNA isolation is performed and the cDNA insert can be subcloned into an appropriate vector for restriction mapping and sequencing. If the phage vector is Lambda ZAP® II, coinfection with helper phage allows rescue and

- 18 -

recircularization of pBluescript SK⁻ phagemid vector (Stratagene, La Jolla, CA) harboring the cDNA; alternatively the phage clone is purified and the cDNA insert is subcloned into a vector suitable for
5 restriction mapping and sequencing. If the clone does not contain the entire hITF gene (as assessed by homology to rITF and the presence of start and stop codons), the library can be rescreened with the original rITF probe or, preferably, with a probe generated from the hITF
10 clone obtained. If none of the clones contain the intact gene, it can be reconstructed from clones which bear overlapping fragments of hITF.

Direct Isolation of an hITF Probe by PCR

It is possible to isolate part of the hITF gene
15 directly from the packaged library or cDNA. To isolate a portion of hITF directly from the packaged library, a pair of oligonucleotide primers and Taq polymerase are used to amplify the DNA corresponding to the hITF gene. The primers used would be approximately 15-20 nucleotides
20 long and correspond in sequence to the 5'-most and 3'-most portions of the rITF coding sequence. Friedman et al. (In PCR Protocols: A Guide to Methods and Applications, Innis et al., Eds., Academic Press, San Diego, CA) describe a procedure for such amplification.
25 Briefly, phage particles are disrupted by heating; Taq polymerase, primers (300 pmol of each), dNTPs, and Taq polymerase buffer are added; and the mixture is thermally cycled to amplify DNA. The amplified DNA is isolated by agarose gel electrophoresis. The ends of the fragment
30 are prepared for ligation into an appropriate vector by making them flush with T4 polymerase and, if desired, adding linkers. Alternatively, a restriction site may be engineered into the fragment by using primers which have sequence added to their 5' ends which sequence will

- 19 -

generate an appropriate sticky end when digested. For example the sequence: 5'-GGGCGGCCGC-3' (SEQ ID NO:4) can be added to the 5' end of each primer. This sequence includes the *NotI* restriction site flanked at the 5' end by the sequence: GG. The additional nucleotides prevent the 5' ends from denaturing and interfering with subsequent restriction digestion with *NotI*. The gel purified DNA of the appropriate size is next cloned into a cloning vector for sequencing and restriction mapping.

10 This clone will not have the entire hITF sequence, rather it will be a combination of hITF (the region between the sequences corresponding to the primers) and rITF (the 5' and 3' ends which correspond to the primer sequences). However, this DNA can be used to generate a labelled

15 probe (produced by nick translation or random primer labelling) which, since it is the correct hITF sequence, can be used in a high stringency screening of the library from which the cDNA was originally isolated. In an alternative approach, cDNA can be used in the above

20 procedure instead of a packaged library. This eliminates the steps of modifying the cDNA for insertion into a vector as well as cDNA packaging and library amplification. Ausubel et al. *supra* provides a protocol for amplification of a particular DNA fragment directly

25 from cDNA and a protocol for amplification from poly(A)⁺ RNA.

Identification of a Presumptive Human ITF clone

A nick translated probe derived from rITF cDNA (corresponding to nucleotides 1 to 431 of SEQ ID NO:1)

30 was used for Northern blot analysis of poly(A)⁺ RNA derived from human intestinal mucosal scrapings. Probe hybridization and blot washing were carried out according to standard procedures. Probe (5×10^5 cpm/ml hybridization buffer) was hybridized to the filter at

- 20 -

45°C in 5X SSC with 30% formamide. The filter was then washed at 60°C in 5X SSC with 40% formamide. Using this protocol, a band was clearly visible after an overnight exposure of the filter with an intensifying screen. This result indicated that there is sufficient homology between rITF and hITF to allow the use of probes derived from the sequence of the rITF gene for identification of the hITF gene.

A human intestinal cDNA library was obtained from Clontech (Palo Alto, CA). Alternatively, a human intestinal cDNA library may be produced from mucosal scrapings as described above. Four oligonucleotide probes were selected for screening the library cDNA. Two of the probes correspond to sequences within the region of rITF encoding the trefoil and are referred to as internal probes (5'-GTACATTCTGTCTCTTGCAGA-3' (SEQ ID NO:5) and 5'-TAACCCTGCTGCTGCTGCTCCTGG-3' (SEQ ID NO:6). The other two probes recognize sequences within rITF but outside of the trefoil encoding region and are referred to as external probes (5'-GTTTGCGTGCTGCCATGGAGA-3' (SEQ ID NO:7) and 5'-CCGCAATTAGAACAGCCTTGT-3' (SEQ ID NO:8). These probes were tested for their utility by using them to screen the rat intestinal cDNA library described above. Each of the four probes could be used to identify a clone harboring all or part of the rITF gene. This result indicates that these probes may be used to screen the human intestinal library for the presence of hITF.

The internal probes were used as described above to amplify a DNA fragment from human colon library cDNA (Clontech, Palo Alto, CA). Linkers were added to the isolated DNA fragment which was then inserted into pBluescript phagemid vector (Stratagene, La Jolla, CA). The region of this clone corresponding to the sequence of human cDNA (i.e., not including the sequence corresponding to the internal probes) was used to make a

- 21 -

radioactively labelled probe by random oligonucleotide-primed synthesis (Ausbel et al., *supra*). This probe was then used to screen the human colon cDNA library. This screening led to the identification of 29 clones. One of these clones (HuPCR-ITF) was nick-translated to generate a probe for Northern analysis of poly(A)⁺ RNA isolated from human intestinal mucosal scrapings. A single band of roughly the same size as the rat transcript (approximately 0.45 kDa) was observed.

10 Northern analysis of poly(A)⁺ isolated from human tissues indicated that RNA corresponding to this probe was expressed in the small intestine and the large intestine but not in the stomach or the liver. These results indicate that the clone does not encode the human
15 homolog of porcine PSP. Porcine PSP is expressed in porcine pancreas and is not significantly expressed in the small or large intestine. These results also distinguish the cloned gene from pS2 which is expressed in the stomach.

20 Figure 6 shows the nucleic acid sequence information for human ITF cDNA, along with the deduced amino acid sequence in one-letter code (SEQ ID NO:3). This clone was obtained by the methods described above.

Production of hITF

25 The isolated hITF gene can be cloned into a mammalian expression vector for protein expression. Appropriate vectors include pMAMneo (Clontech, Palo Alto, CA) which provides a RSV-LTR enhancer linked to a dexamethasone-inducible MMTV-LTR promoter, an SV40 origin
30 of replication (allows replication in COS cells), a neomycin gene, and SV40 splicing and polyadenylation sites. This vector can be used to express the protein in COS cells, CHO cells, or mouse fibroblasts. The gene may

- 22 -

also be cloned into a vector for expression in drosophila cells using the baculovirus expression system.

Purification of Intestinal Trefoil Factor

Intestinal trefoil factor can be purified from
5 intestinal mucosal scrapings of human, rats or any other species which expresses ITF (pigs and cows may provide a source of ITF). The purification procedure used for PSP will be useful for the purification of ITF since the proteins are likely to be homologous. Jorgensen et al.
10 describes a method for purification of PSP (*Regulatory Peptides* 3:207, 1982). The preferred method is the second approach described by Jorgensen et al. (*supra*). This method involves chromatography of SP-Sephadex C-25 and QAE Sephadex A-25 columns (Sigma, St. Louis, MO) in
15 acidic buffer.

Anti-Intestinal Trefoil Factor Monoclonal Antibodies

Anti-intestinal trefoil factor monoclonal
antibodies can be raised against synthetic peptides whose
20 sequences are based on the deduced amino acid sequence of cloned hITF (SEQ ID NO:3). Most commonly the peptide is based on the amino-or carboxy-terminal 10-20 amino acids of the protein of interest (here, hITF). The peptide is usually chemically cross-linked to a carrier molecule
25 such as bovine serum albumin or keyhole limpet hemocyanin. The peptide is selected with the goal of generating antibodies which will cross-react with the native hITF. Accordingly, the peptide should correspond to an antigenic region of the peptide of interest. This
30 is accomplished by choosing a region of the protein which is (1) surface exposed, e.g., a hydrophobic region or (2) relatively flexible, e.g., a loop region or a β -turn region. In any case, if the peptide is to be coupled to

- 23 -

a carrier, it must have an amino acid with a side chain capable of participating in the coupling reaction. See Hopp et al. (*Mol. Immunol.* 20:483, 1983; *J. Mol. Biol.* 157:105, 1982) for a discussion of the issues involved in the selection of antigenic peptides. A second consideration is the presence of a protein homologous to hITF in the animal to be immunized. If such a protein exists, it is important to select a region of hITF which is not highly homologous to that homolog.

For hITF, peptides that correspond to the amino-terminal or carboxy-terminal 15 amino acids are likely to be less homologous across species and exposed to the surface (and thus antigenic). Thus they are preferred for the production of monoclonal antibodies. Purified hITF can also be used for the generation of antibodies.

Genetic Disruption of a Trefoil Protein Impairs the

Defense of Intestinal Mucosa

As stated above, ITF is a member of the family of trefoil proteins that are expressed specifically and abundantly at the mucosal surface of the gastrointestinal tract. Other members of this family include pS2, which is expressed almost exclusively by foveolar cells of the stomach (Masiakowski et al., *Nucl. Acids. Res.* 10:7896, 1982; Jorgensen et al., *Regulatory Peptides* 3:231, 1982), and pancreatic spasmolytic peptide (SP), which is expressed by the pancreas and by gastric antrum (Jorgensen et al., *supra*). As described above, the expression of these proteins is enhanced in the proximity of the injured bowel. In order to study the role of ITF in vivo, the gene was rendered non-functional by targeted disruption in mice.

- 24 -

Isolation of the Murine ITF Gene and Generation of ITF-Deficient Mice

The murine *ITF* gene was isolated from a phage genomic library using the rat *ITF* cDNA sequence as a probe, and its identity was confirmed by nucleotide sequencing using standard techniques (Mashimo et al., *Biochem. Biophys. Res. Comm.* 210:31, 1995).

A targeting vector for disrupting the gene by homologous recombination in embryonic stem (ES) cells was designed and constructed, as shown in Figure 7. The entire second exon (Ex2) of the murine *ITF* gene, which is contained within the *XbaI*-*EcoRI* fragment shown, was replaced with the neomycin resistance (*neo*) gene cassette. As the deleted sequence encodes most of the "trefoil domain," the ability of any resultant peptides to produce the looping structure characteristic of trefoil proteins is abolished. A positive-negative selection strategy (Mansour et al., *Nature* 336:348, 1988) was used to enrich for homologous recombination events in the embryonic stem (ES) cells by selecting for *neo* within the homologous DNA and against a herpes simplex virus thymidine kinase gene (*hsv-tk*) placed at the 3' end of the targeting vector. The pPNT plasmid (Tybulewicz et al., *Cell* 65:1153, 1991) was used to construct the targeting vector. The targeting vector was linearized with the restriction enzyme *NotI* and electroporated into pluripotent J1 ES cells (Li et al., *Cell* 69:915, 1992) under conditions previously described (Strittmatter et al., *Cell* 80:445, 1995). Disruption of the *ITF* gene in ES cells following homologous recombination was distinguished from random integration of the targeting vector by Southern blot analysis of genomic DNA from individual clones of cells digested with the restriction enzyme *XhoI*. The pITF2 probe identified a 19 kb "wild type" fragment and a 23 kb "knock out" fragment created

- 25 -

by introduction of an XhoI site upon homologous insertion of the targeting vector. Approximately 10% of neomycin-resistant ES clones were found to have undergone homologous ITF recombination using this method.

- 5 The polymerase chain reaction (PCR) was used to confirm the targeted mutation as follows. A 200 bp region of DNA was amplified using primers spanning exon 2 of *ITF* (5'-GCAGTGTAACAACCGTGGTTGCTGC-3' (SEQ ID NO:9) and 5'-TGACCCTGTGTCATCACCCTGGC-3' (SEQ ID NO:10)); and a 400
10 bp region of the *neo* gene was amplified with a second set of primers (5'-CGGCTGCTCTGATGGCCGCC-3' (SEQ ID NO:11) and 5'-GCCGGCCACAGTCGATGAATC-3' (SEQ ID NO:12)). The DNA template for the PCR reaction was obtained from tail
15 animal, and the samples were digested with proteinase-K (200 μ l at 0.5 mg/ml in 50 mM Tris-HCl pH 8.0 and 0.5% Triton X-100; Sigma, St. Louis, MO) at 55°C overnight. One μ l of this mixture was added directly to a 25 μ l PCR reaction (per Stratagene, Menosha, WI). The reaction was
20 begun with a "hot start" (incubation at 96°C for 10 minutes), and the following cycle was repeated 30 times: 72°C for 120 seconds (hybridization and elongation) and 96°C for 30 seconds (denaturation). Ten μ l of each reaction mixture was electrophoresed on a 2% agarose gel.
25 Wild type animals were identified by the presence of a 200 bp fragment, corresponding to an intact *ITF* gene, heterozygous animals were identified by the presence of this band and, in addition, a 400 bp fragment produced by amplification of the *neo* gene, and *ITF*-deficient (knock
30 out) animals were identified by the presence of only the fragment corresponding to the *neo* gene.

Two ES clones, which arose independently, were used to derive two lines of mice lacking *ITF*. These mice were screened by Southern genomic blot analysis as
35 described for ES clones, or by PCR.

- 26 -

Analysis of Trefoil Peptide Expression in
Wild Type and Mutant Mice

Although expression of ITF is abolished in the mutant mice, expression of other trefoil genes is preserved. Northern blot analysis was performed using cDNA probes for ITF (Suemori et al., *Proc. Natl. Acad. Sci. USA* **88**:11017, 1991), SP (Jeffrey et al. *Gastroenterology* **106**:336, 1994), and, as a positive control, glyceraldehyde 3-phosphate dehydrogenase (GAPDH). The nucleic acid probe for murine pS2 was made by reverse transcription-polymerase chain reaction (RT-PCR) using the oligonucleotide pairs:
5'-GAGAGGTTGCTGTTTGTATGACA-3' (SEQ ID NO:13) and
5'-GCCAAGTCTTGATGTAGCCAGTT-3" (SEQ ID NO:14), which were synthesized based on the published mouse pS2 cDNA sequence (GenBank Accession Number: Z21858). The GeneAmp RNA PCR Kit (Perkin Elmer) was used according to the manufacturer's instructions, as was the pCR^{II} (Invitrogen) cloning vector. RNA was extracted from the following tissues from both wild type and ITF-deficient (knock out) mice: stomach, duodenum, terminal ileum, right colon, appendix, transverse colon, left colon, and rectum. Fifteen μ g of total RNA from each sample were electrophoresed on a 1% agarose gel, and transferred to nitrocellulose paper. Following hybridization, washing, and autoradiography, wild type mice exhibited a pattern of tissue expression considered normal: ITF was expressed in the small intestine and colon, which is the same expression pattern seen for ITF in the rat and human. The analysis of mutant mice confirmed the lack of ITF expression in the gastrointestinal tract. In contrast, the expression of the other trefoil proteins, SP and pS2, are unaltered in the gastrointestinal tract of mutant mice. SP was expressed in the stomach and, at lower levels, in the duodenum of both wild type and

- 27 -

mutant mice. Similarly, pS2 was expressed in the stomach of both wild type and ITF-deficient mice.

Immunocytochemistry Reveals that ITF is not
Expressed in the Colon of ITF-deficient mice

5 In order to confirm that ITF protein was not expressed by ITF knock out mice, immunocytochemistry was performed as follows. Tissue from the colon and small intestine was fixed in the course of perfusion, immersed in 4% paraformaldehyde (McLean et al., *J. Histochem.*
10 *Cytochem.* 22:1077, 1974), and embedded in paraffin. Sections were collected and stained either with a polyclonal antibody raised against a synthetic peptide from the predicted 18 carboxy-terminal amino acids of murine ITF or a monoclonal antibody against colonic mucin
15 (Podolsky et al., *J. Clin. Invest.* 77:1263, 1986). Primary antibody binding was visualized with a biotinylated secondary antibody, Avidin DH, biotinylated horseradish peroxidase H, and diaminobenzidine tetrahydrochloride reagents according to the
20 manufacturer's instructions (VectaStain ABC, Vector Laboratories, Burlingame, CA). Following immunocytochemistry, the sections were counterstained with hematoxylin and viewed. Goblet cells in the colon of wild type mice were immunoreactive with both
25 antibodies, staining positively for ITF and mucin. In contrast, the goblet cells in the colon of ITF-deficient mice lacked detectable ITF but continued to express colonic mucin.

Induction of Mild Colonic Epithelial Injury
with Dextran Sulfate Sodium

30 ITF-deficient mice derived from each ES clone appear to develop normally and are grossly indistinguishable from heterozygous and wild type litter

- 28 -

mates. Their growth is not retarded and they reach maturity without evident diarrhea or occult fecal blood loss. However, the colon of ITF-deficient mice may be more prone to injury than the colon of wild type mice.

5 To investigate this hypothesis, dextran sulfate sodium (DSS), which reproducibly creates mild colonic epithelial injury with ulceration in mice (Kim et al., *Scand. J. Gastroent.* 27:529, 1992; Wells et al., *J. Acquired Immune Deficiency Syndrome* 3:361, 1990; Okayasu et al.,

10 *Gastroenterology* 98:694, 1990) was administered in the animals' drinking water. After standardization of DSS effects in comparable wild type mice, a group of 20 wild type and 20 ITF-deficient mice (litter mates from heterozygous crosses, weighing > 20 grams each) were

15 treated with 2.5% DSS in their drinking water for nine days.

Although 85% of wild type mice and 100% of ITF-deficient mice treated with DSS demonstrate occult blood (using Hemocult, Smith Kline Diagnostics, San Jose, CA)

20 in their stool during the period of treatment, ITF-deficient mice were markedly more sensitive to the injurious effects of DSS. Fifty percent of ITF-deficient mice developed frankly bloody diarrhea and died (Fig. 8). In contrast, only 10% of wild type mice treated similarly

25 exhibited bloody diarrhea, and only 5% died. Weight loss was also significantly more pronounced in ITF-deficient mice than wild type mice receiving DSS.

ITF-Deficient Mice Treated with Dextran Sulfate Sodium (DSS) Develop Severe Colonic Erosions

30 After seven days of treatment with DSS (2.5% w/v), the colons of wild type and ITF-deficient mice were examined histologically. Left colon transections were fixed in 4% paraformaldehyde, mounted in paraffin, and stained with hematoxylin and eosin. Multiple sites of

- 29 -

obvious ulceration and hemorrhage were present in the colon of ITF-deficient mice, while the colons of most wild type mice were grossly indistinguishable from those of untreated mice. Histological examination of the DSS-
5 treated ITF-deficient colon confirmed the presence of multiple erosions and intense inflammatory changes including crypt abscesses. Damage was more pronounced in the distal colon, i.e., the descending colon, sigmoid colon, and rectum, which contained large, broad areas of
10 mucosal ulceration. When similarly inspected, mucosal erosions could be seen in the tissue of 80% of the DSS-treated wild type mice, but most were small lesions that also appeared to be healing, with complete re-epithelialization of most lesions. There was no evidence
15 of re-epithelialization in the colons of ITF-deficient mice exposed to DSS.

During the normal course of growth and development, intestinal epithelial cells originate from stem cells in the intestinal crypts and rapidly progress
20 up the crypt and villus to be extruded from the villus tip within five days. After intestinal injury, the epithelial covering is repopulated by cells which appear to generate signals to heal the lesion by modulation of epithelial and mesenchymal cell growth and matrix
25 formation (Poulsom et al., *J. Clin. Gastroenterol.* 17:S78, 1993). *In vitro* evidence suggests that trefoil proteins play a key role in re-establishing mucosal integrity after injury. Despite the normal restriction of SP and pS2 expression to the proximal gastrointestinal
30 tract, these trefoil proteins and ITF are abundantly expressed at sites of colonic injury and repair. The DSS model described above provides a system for testing the protective effects of ITF, other trefoil peptides, or active polypeptide fragments or variants thereof. One
35 can administer a molecule to be tested to DSS-treated

- 30 -

mice, either wild type or ITF-deficient mice, and determine whether the molecule has therapeutic effects by performing the assays described above.

In addition to the use of DSS, any chemical compound that is known to damage the mucosa lining the digestive tract can be used to assay the proteins of the invention. These compounds include, but are not limited to, alcohol, indomethacin, and methotrexate. For example, methotrexate (MTX) can be administered intraperitoneally to mice at a dose of 40 mg/kg. One group of MTX-treated animals could be given, in addition, the protein in question. Various parameters, such as body weight, the presence of lesions in the digestive tract, and mortality of these animals could then be compared to equivalent measurements taken from animals that were not treated with the protein.

In Situ *H. pylori* Binding Assay

One method for determining whether a given protein (or protein fragment or variant) is useful in the prevention or treatment of diseases associated with *H. pylori* infection is to examine it in the context of an established animal model of *H. pylori* infection. One such model was recently developed by Falk et al. (*Proc. Natl. Acad. Sci. USA* 92:1515-1519, 1995). This model involves the use of transgenic mice that express the enzyme α -1,3/4-fucosyltransferase and, as a consequence, express Le^b on the surface of mucosal cells that bound clinical isolates of *H. pylori*. If the addition of a protein, such as ITF, to this system reduces the level of *H. pylori* binding to the mucosal cell, the protein would be considered an inhibitor of *H. pylori*. More specifically, the assay could be carried out as follows. *H. pylori* are obtained, for example, from patients with gastric ulcers or chronic active gastritis, grown to

- 31 -

stationary phase, and labeled, for example with digoxigenin or fluorescein isothiocyanate (FITC). The labeled bacteria are then exposed, together with the protein of interest, to frozen sections prepared from the stomach, duodenum, ileum, or liver of adult transgenic mice (as described above). As a control, the experiment could be performed in parallel using tissue from a wild type littermate. The sections are fixed with ice-cold methanol for 5 minutes, rinsed three times with wash buffer (TBS; 0.1 mM CaCl_2 , 1 mM MnCl_2 , 1 mM MgCl_2 ; 10 minutes/cycle), and treated with blocking buffer (Boehringer Mannheim; see also Falk *supra*). Bacteria are diluted to an OD_{600} of 0.05 with dilution buffer [TBS; 0.1 mM CaCl_2 , 1 mM MnCl_2 , 1 mM MgCl_2 containing leupeptin (1 $\mu\text{g}/\text{ml}$), aprotinin (1 $\mu\text{g}/\text{ml}$), [-1-p-tosylamido-2-phenylethyl chloromethyl ketone (100 $\mu\text{g}/\text{ml}$), phenylmethylsulfonyl fluoride (100 $\mu\text{g}/\text{ml}$), and pepstatin A (1 $\mu\text{g}/\text{ml}$)] and overlaid on the sections for 2 hours at room temperature in a humidified chamber. Slides are then washed six times in wash buffer on a rotating platform (5 minutes/cycle at room temperature). Digoxigenin-labeled bacteria are visualized on washed slides with FITC-conjugated sheep anti-digoxigenin immunoglobulin (Boehringer Mannheim) diluted 1:100 in histoblocking buffer. Nuclei were stained with bisbenzimidazole (Sigma). For blocking controls, digoxigenin-conjugated stationary-phase bacteria can be suspended in dilution buffer to an OD_{600} of 0.05 and shaken with or without Le^b -HSA or Le^a -HSA (final concentration, 50 $\mu\text{g}/\text{ml}$; reaction mixture, 200 μl) for 1 hour at room temperature. The suspension is then overlaid on methanol-fixed frozen sections.

- 32 -

Use

In the practice of the present invention, ITF may be administered as described below for treatment of peptic ulcer diseases, inflammatory bowel diseases, for protection of the intestinal tract from injury caused by bacterial infection, radiation injury, or other insults. Tissues that are not a part of the alimentary canal can also be treated. These tissues include the skin, the corneal surface of the eye, and tissues within the respiratory and genitourinary tract. The mode of administration, dosage, and formulation of ITF will depend upon the condition being treated. Further guidance regarding treatment regimes is given below. Furthermore, treatment may begin before an injury has occurred because the polypeptides and compositions of the invention are believed to exert a protective effect.

Other EmbodimentsProduction of Antibodies

ITF may be used to produce monoclonal antibodies for the detection of ITF in intestinal tissue or blood serum by means of an indirect immunoassay. ITF may be detectably labelled and used in an *in situ* hybridization assay for the detection of ITF binding sites. Labels may include, but are not limited to, fluorescein or a radioactive ligand.

ITF may be used to protect and stabilize other proteins. This protection is accomplished by forming a hybrid molecule in which all or part of ITF is fused to either the carboxy-terminus or the amino-terminus (or both) of the protein of interest. Because ITF is resistant to degradation in the digestive system, it will protect the protein of interest from such degradation. As a consequence, the protein of interest is likely to

- 33 -

remain active in the digestive system and/or will be more readily absorbed in an intact form.

Stably dimerized trefoil protein can be used in the methods of the invention. Such molecules can be prepared by stably crosslinking monomers of trefoil or by expressing a gene encoding a tandem repeat of a trefoil protein (e.g., ITF) or a portion thereof (e.g., a portion capable of forming the three loop structure characteristic of trefoil proteins). Also useful in the method of the invention are trefoil proteins produced by chemical synthesis.

The invention also encompasses antibodies that bind the polypeptides of the invention, i.e., trefoil polypeptides such as ITF. Antibodies that specifically recognize one or more epitopes of these polypeptides, or fragments thereof, are also encompassed by the invention. Such antibodies include but are not limited to polyclonal antibodies, monoclonal antibodies (mAbs), humanized or chimeric antibodies, single chain antibodies, Fab fragments, F(ab')₂ fragments, fragments produced by a Fab expression library, anti-idiotypic (anti-Id) antibodies, and epitope-binding fragments of any of the above.

The antibodies of the invention may be used, for example, in the detection of ITF in a biological sample and may, therefore, be utilized as part of a diagnostic or prognostic technique whereby patients may be tested for abnormal amounts of ITF. Typically, the expression of ITF is downregulated in the immediate vicinity of a lesion, such as that caused by inflammatory bowel diseases (for example, colitis).

For the production of antibodies, various host animals may be immunized by injection with a peptide having a sequence that is present, for example, in ITF. Such host animals may include but are not limited to rabbits, mice, and rats, to name but a few. Various

- 34 -

adjuvants may be used to increase the immunological response, depending on the host species, including but not limited to Freund's (complete and incomplete), mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanin, dinitrophenol, and potentially useful human adjuvants such as BCG (bacille Calmette-Guerin) and *Corynebacterium parvum*. Polyclonal antibodies are heterogeneous populations of antibody molecules derived from the sera of the immunized animals.

Monoclonal antibodies, which are homogeneous populations of antibodies to a particular antigen, may be obtained by any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique of Kohler and Milstein (*Nature* 256:495-497, 1975; and U.S. Patent No. 4,376,110), the human B cell hybridoma technique (Kosbor et al., *Immunology Today* 4:72, 1983; Cole et al., *Proc. Natl. Acad. Sci. USA* 80:2026-2030, 1983), and the EBV-hybridoma technique (Cole et al., "Monoclonal Antibodies And Cancer Therapy," Alan R. Liss, Inc., pp. 77-96, 1985). Such antibodies may be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing the mAb of this invention may be cultivated *in vitro* or *in vivo*. Production of high titers of mAbs *in vivo* makes this the presently preferred method of production.

In addition, techniques developed for the production of "chimeric antibodies" (Morrison et al., *Proc. Natl. Acad. Sci. USA*, 81:6851-6855, 1984; Neuberger et al., *Nature*, 312:604-608, 1984; Takeda et al., *Nature*, 314:452-454, 1985) by splicing the genes from a mouse antibody molecule of appropriate antigen specificity

- 35 -

together with genes from a human antibody molecule of appropriate biological activity can be used. A chimeric antibody is a molecule in which different portions are derived from different animal species, such as those
5 having a variable region derived from a murine mAb and a human immunoglobulin constant region.

Alternatively, techniques described for the production of single chain antibodies (U.S. Patent 4,946,778; Bird, *Science* 242:423-426, 1988; Huston et
10 al., *Proc. Natl. Acad. Sci. USA* 85:5879-5883, 1988; and Ward et al., 1989, *Nature* 334:544-546, 1989) can be adapted to produce single chain antibodies against trefoil polypeptides such as ITF. Single chain
15 antibodies are formed by linking the heavy and light chain fragments of the Fv region via an amino acid bridge, resulting in a single chain polypeptide.

Antibody fragments which recognize specific epitopes may be generated by known techniques. For example, such fragments include but are not limited to:
20 the F(ab')₂ fragments which can be produced by pepsin digestion of the antibody molecule and the Fab fragments which can be generated by reducing the disulfide bridges of the F(ab')₂ fragments. Alternatively, Fab expression libraries may be constructed (Huse et al., *Science*,
25 246:1275-1281, 1989) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

These antibodies can, in turn, be utilized to generate anti-idiotypic antibodies that "mimic" ITF, using
30 techniques well known to those skilled in the art. (See, for example, Greenspan and Bona, *FASEB J.* 7:437-444, 1993; and Nissinoff, *J. Immunol.* 147:2429-2438, 1991). Such neutralizing anti-idiotypes or Fab fragments of such anti-idiotypes can be used in diagnostic regimens to
35 detect disorders associated with apoptotic cell death.

- 36 -

Antibodies can be humanized by methods known in the art. For example, monoclonal antibodies with a desired binding specificity can be commercially humanized (Scotgene, Scotland; Oxford Molecular, Palo Alto, CA). Fully human antibodies, such as those expressed in transgenic animals are also features of the invention (Green et al., *Nature Genetics* 7:13-21, 1994; see also U.S. Patents 5,545,806 and 5,569,825, both of which are hereby incorporated by reference).

10 Administration of ITF to Protect or Treat Tissues
 Outside the Alimentary Canal

The polypeptides of the invention, including ITF, analogs, and fragments thereof, as well as other trefoil factors, such as SP (spasmolytic polypeptide) and PS2, can be used to protect or treat tissues that are not found within the alimentary canal. The polypeptides can be used, for example, to treat any sort of wound, such as a lesion, an ulcer, a burn, or an abrasion on the skin, the surface of the eye (i.e., the cornea), or within the respiratory or genitourinary tracts. The exact nature of the injury and the cause of the injury need not be precisely defined.

Regardless of the location of the injury (i.e., regardless of whether or not the injury is within the alimentary canal), toxicity and therapeutic efficacy of a given compound can be determined by standard pharmaceutical procedures, using either cells in culture or experimental animals to determine the LD₅₀ (the dose lethal to 50% of the population) and the ED₅₀ (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD₅₀/ED₅₀. Compounds which exhibit large therapeutic indices are preferred. While compounds that exhibit

- 37 -

toxic side effects may be used, care should be taken to design a delivery system that targets such compounds to the site of affected tissue in order to minimize potential damage to unaffected cells and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in humans. The dosage of such compounds lies preferably within a range of circulating concentrations that include the ED_{50} with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For any compound used in the method of the invention, the therapeutically effective dose can be estimated initially from cell culture assays. A dose may be formulated in animal models to achieve a circulating plasma concentration range that includes the IC_{50} (that is, the concentration of the test compound which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma may be measured, for example, by high performance liquid chromatography.

Pharmaceutical compositions for use in accordance with the present invention may be formulated in conventional manner using one or more physiologically acceptable carriers or excipients. The pharmaceutical compositions can also contain mucin glycoproteins.

Thus, the compounds and their physiologically acceptable salts and solvates may be formulated for administration by inhalation or insufflation (either through the mouth or the nose) or oral, buccal, parenteral, or rectal administration.

As trefoil polypeptides are not degraded within the digestive tract, it is expected that the route of

- 38 -

administration will be oral. The polypeptide could be administered, for example, in the form of a tablet, capsule, or pill, or could be suspended in a solution, such as a syrup, that the patient swallows.

5 Alternatively, the solution containing the polypeptide may be administered as a gastric lavage. The polypeptide may also be included in a solution that is administered as an enema, or it may be administered as a suppository.

For oral administration, which can be used to
10 treat injured tissue within the alimentary canal, the pharmaceutical compositions may take the form of, for example, tablets or capsules prepared by conventional means with pharmaceutically acceptable excipients such as binding agents (for example, pregelatinised maize starch,
15 polyvinylpyrrolidone or hydroxypropyl methylcellulose); fillers (for example, lactose, microcrystalline cellulose or calcium hydrogen phosphate); lubricants (for example, magnesium stearate, talc or silica); disintegrants (for example, potato starch or sodium starch glycolate); or
20 wetting agents (for example, sodium lauryl sulphate). The tablets may be coated by methods well known in the art. Liquid preparations for oral administration may take the form of, for example, solutions, syrups or suspensions, or they may be presented as a dry product
25 for constitution with water or other suitable vehicle before use. Such liquid preparations may be prepared by conventional means with pharmaceutically acceptable additives such as suspending agents (for example, sorbitol syrup, cellulose derivatives or hydrogenated
30 edible fats); emulsifying agents (for example, lecithin or acacia); non-aqueous vehicles (for example, almond oil, oily esters, ethyl alcohol or fractionated vegetable oils); and preservatives (for example, methyl or propyl-p-hydroxybenzoates or sorbic acid). The preparations may
35 also contain buffer salts, flavoring, coloring and

- 39 -

sweetening agents as appropriate. Preparations for oral administration may be suitably formulated to give controlled release of the active compound.

For buccal administration, which can be used to
5 treat injured tissue within the mouth, throat, or upper esophagus, the compositions may take the form of tablets or lozenges formulated in a conventional manner.

For administration by inhalation, which can be used to treated injured tissue within the respiratory
10 tract, the compounds for use according to the present invention are conveniently delivered in the form of an aerosol spray presentation from pressurized packs or a nebulizer, with the use of a suitable propellant, for example, dichlorodifluoromethane, trichlorofluoromethane,
15 dichlorotetrafluoroethane, carbon dioxide or other suitable gas. In the case of a pressurized aerosol the dosage unit may be determined by providing a valve to deliver a metered amount. Capsules and cartridges of, for example, gelatin for use in an inhaler or insufflator
20 may be formulated containing a powder mix of the compound and a suitable powder base such as lactose or starch.

Compositions containing the polypeptides of the invention can also be formulated for parenteral administration by injection, for example, by bolus
25 injection or continuous infusion. Formulations for injection may be presented in unit dosage form, for example, in ampoules or in multi-dose containers, with an added preservative. The compositions may take such forms as suspensions, solutions or emulsions in oily or aqueous
30 vehicles, and may contain formulatory agents such as suspending, stabilizing and/or dispersing agents. Alternatively, the active ingredient may be in powder form for constitution with a suitable vehicle, for example, sterile pyrogen-free water, before use.

- 40 -

The compositions can also be formulated in rectal compositions such as suppositories or retention enemas, for example, containing conventional suppository bases such as cocoa butter or other glycerides.

5 In addition to the formulations described previously, the compounds may also be formulated as a depot preparation. Such long acting formulations may be administered by implantation (for example subcutaneously or intramuscularly) or by intramuscular injection. Thus,
10 for example, the compounds may be formulated with suitable polymeric or hydrophobic materials (for example as an emulsion in an acceptable oil) or ion exchange resins, or as sparingly soluble derivatives, for example, as a sparingly soluble salt.

15 The compositions may, if desired, be presented in a pack or dispenser device which may contain one or more unit dosage forms containing the active ingredient. The pack may for example comprise metal or plastic foil, such as a blister pack. The pack or dispenser device may be
20 accompanied by instructions for administration.

The therapeutic compositions of the invention can also contain a carrier or excipient, many of which are known to skilled artisans. Excipients which can be used include buffers (for example, citrate buffer, phosphate
25 buffer, acetate buffer, and bicarbonate buffer), amino acids, urea, alcohols, ascorbic acid, phospholipids, proteins (for example, serum albumin), EDTA, sodium chloride, liposomes, mannitol, sorbitol, and glycerol. The nucleic acids, polypeptides, antibodies, or
30 modulatory compounds of the invention can be administered by any standard route of administration. For example, administration can be parenteral, intravenous, subcutaneous, intramuscular, intracranial, intraorbital, ophthalmic, intraventricular, intracapsular, intraspinal,
35 intracisternal, intraperitoneal, transmucosal, or oral.

- 41 -

The modulatory compound can be formulated in various ways, according to the corresponding route of administration. For example, liquid solutions can be made for ingestion or injection; gels or powders can be made for ingestion, inhalation, or topical application. Methods for making such formulations are well known and can be found in, for example, "Remington's Pharmaceutical Sciences. It is expected that the preferred route of administration will be oral.

10 It is well known in the medical arts that dosages for any one patient depend on many factors, including the general health, sex, weight, body surface area, and age of the patient, as well as the particular compound to be administered, the time and route of administration, and
15 other drugs being administered concurrently.

Dosages for the polypeptides and antibodies of the invention will vary. For oral administration, the polypeptide can be administered in dosages of about 10 mg to about 500 mg. For example 10, 50, 100, 200, 250, 300,
20 400, or 500 mg can be administered. These dosages can be administered on a periodic basis. For example, a dose may be taken one to four times per day. For topical administration, the polypeptide can be administered in dosages of about 1 to about 10 mg/ml within an ointment,
25 or cream. This composition can also be administered periodically, if necessary. For other routes of administration, dosage will also vary, for example, from about 0.1 to 1,000 mg per application. Determination of the correct dosage within a given therapeutic regime is
30 well within the abilities of one of ordinary skill in the art of pharmacology.

In order to determine the efficacy of a polypeptide in treating a particular disorder, those of skill in the art can perform routine studies using any
35 one of several well known models of injury. For example,

- 42 -

the efficacy of a polypeptide in treating injuries to the cornea can be carried out using the *in vitro* model for corneal wound healing described by Collin et al. (*Current Eye Res.* 14:331-339, 1995). In this model system, 5 corneas that were unsuitable for transplantation were obtained from human organ donors and used within 5 days post-mortem. A cautery tip was used to create a linear non-perforating thermal burn approximately 5 mm in length on the cornea. The wounded corneas were immediately 10 dissected and placed in an air/liquid organ culture system (as described in Richard et al., *Curr. Eye Res.* 10:739-749, 1991; and Anderson et al., *Ophthalmol. Vis. Sci.* 3:442-449, 1993). Thus, to determine the efficacy of a polypeptide of the invention in treating such an 15 injury, one would simply apply the polypeptide to the wounded cornea, for example, by placing the polypeptide in the tissue culture medium, and assessing the effect of the polypeptide on wound healing in injured, versus uninjured, corneas. If additional guidance is required 20 in assessing the wound, skilled artisans may again consult Collin et al. (*supra*), who also describes histochemical analysis of the wounded corneas. Protocols for a wound closure model using cultured rabbit corneal endothelial cells can also be used (for example, see 25 Joyce et al., *Invest. Ophthalmol. Vis. Sci.* 31:1816-1826, 1990). Alternatively, to assess the efficacy of a polypeptide in the context of a physical wound to the cornea, the injury induced as described by Kessler (*Curr. Eye Res.* 14:985-992, 1995) can be used. Similarly, 30 numerous models are available in which to test the efficacy of a polypeptide in preventing or healing a wound to the epidermis. Persons of ordinary skill in the art are well aware of these models and able to carry out the procedures described in the art without resort to 35 undue experimentation.

- 43 -

Other embodiments are within the following claims.

- 44 -

SEQUENCE LISTING

(1) GENERAL INFORMATION

- (i) APPLICANT: The General Hospital Corporation
- (ii) TITLE OF THE INVENTION: INTESTINAL TREFOIL PROTEINS
- 5 (iii) NUMBER OF SEQUENCES: 18
- (iv) CORRESPONDENCE ADDRESS:
- (A) ADDRESSEE: Fish & Richardson, P.C.
- (B) STREET: 225 Franklin Street
- 10 (C) CITY: Boston
- (D) STATE: MA
- (E) COUNTRY: US
- (F) ZIP: 02110-2804
- (v) COMPUTER READABLE FORM:
- (A) MEDIUM TYPE: Diskette
- 15 (B) COMPUTER: IBM Compatible
- (C) OPERATING SYSTEM: Windows95
- (D) SOFTWARE: FastSEQ for Windows Version 2.0
- (vi) CURRENT APPLICATION DATA:
- (A) APPLICATION NUMBER: PCT/US97/----
- 20 (B) FILING DATE: 11-APR-1997
- (C) CLASSIFICATION:
- (vii) PRIOR APPLICATION DATA:
- (A) APPLICATION NUMBER: 08/631,469
- (B) FILING DATE: 12-APR-1996
- 25 (A) APPLICATION NUMBER: 08/191,352
- (B) FILING DATE: 02-FEB-1994
- (A) APPLICATION NUMBER: 08/037,741
- (B) FILING DATE: 25-MAR-1993
- 30 (A) APPLICATION NUMBER: 07/837,192
- (B) FILING DATE: 13-FEB-1992
- (A) APPLICATION NUMBER: 07/655,965
- (B) FILING DATE: 14-FEB-1991
- (viii) ATTORNEY/AGENT INFORMATION:
- (A) NAME: Meiklejohn, Ph.D., Anita L.
- 35 (B) REGISTRATION NUMBER: 35,283
- (C) REFERENCE/DOCKET NUMBER: 00786/322WO1
- (ix) TELECOMMUNICATION INFORMATION:
- (A) TELEPHONE: 617-542-5070
- (B) TELEFAX: 617-542-8906
- 40 (C) TELEX: 200154
- (2) INFORMATION FOR SEQ ID NO:1:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 431 base pairs
- (B) TYPE: nucleic acid
- 45 (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

- 45 -

(ii) MOLECULE TYPE: genomic DNA

(ix) FEATURE:

(A) NAME/KEY: Coding Sequence

(B) LOCATION: 18...260

(D) OTHER INFORMATION:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

GAAGTTTGCG TGCTGCC ATG GAG ACC AGA GCC TTC TGG ATA ACC CTG CTG 50
Met Glu Thr Arg Ala Phe Trp Ile Thr Leu Leu
1 5 10

CTG GTC CTG GTT GCT GGG TCC TCC TGC AAA GCC CAG GAA TTT GTT GGC 98
Leu Val Leu Val Ala Gly Ser Ser Cys Lys Ala Gln Glu Phe Val Gly
15 20 25

CTA TCT CCA AGC CAA TGT ATG GCG CCA ACA AAT GTC AGG GTG GAC TGT 146
Leu Ser Pro Ser Gln Cys Met Ala Pro Thr Asn Val Arg Val Asp Cys
15 30 35 40

AAC TAC CCC ACT GTC ACA TCA GAG CAG TGT AAC AAC CGT GGT TGC TGT 194
Asn Tyr Pro Thr Val Thr Ser Glu Gln Cys Asn Asn Arg Gly Cys Cys
45 50 55

TTT GAC TCC AGC ATC CCA AAT GTG CCC TGG TGC TTC AAA CCT CTG CAA 242
Phe Asp Ser Ser Ile Pro Asn Val Pro Trp Cys Phe Lys Pro Leu Gln
60 65 70 75

GAG ACA GAA TGT ACA TTT TGAAGCTGTC CAGGCTCCAG GAAGGGAGCT CCACACCC 298
Glu Thr Glu Cys Thr Phe
80

TGGACTCTTG CTGATGGTAG TGGCCCAGGG TAACACTCAC CCCTGATCTG CTCCCTCGCG 358

CCGGCCAATA TAGGAGCTGG GAGTCCAGAA GAATAAAGAC CTTACAGTCA GCACAAGGCT 418

GTTCTAATTG CGG 431

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 81 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(v) FRAGMENT TYPE:

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

Met Glu Thr Arg Ala Phe Trp Ile Thr Leu Leu Val Leu Val Ala
1 5 10 15

Gly Ser Ser Cys Lys Ala Gln Glu Phe Val Gly Leu Ser Pro Ser Gln
20 25 30

Cys Met Ala Pro Thr Asn Val Arg Val Asp Cys Asn Tyr Pro Thr Val
35 40 45

Thr Ser Glu Gln Cys Asn Asn Arg Gly Cys Cys Phe Asp Ser Ser Ile
50 55 60

- 46 -

Pro Asn Val Pro Trp Cys Phe Lys Pro Leu Gln Glu Thr Glu Cys Thr
 65 70 75 80

Phe

(2) INFORMATION FOR SEQ ID NO:3:

- 5 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 403 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

- 10 (ii) MOLECULE TYPE: genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

GATGCTGGGG CTGGTCCTGG CCTTGCTGTC CTCCAGCTCT GCTGAGGAGT ACGTGGGCCT 60
 GTCTGCAAAC CAGTGTGCCG TGCCGGCCAA GGACAGGGTG GACTGCGGCT ACCCCCATGT 120
 CACCCCCAAG GAGTGCAACA ACCGGGGGCTG CTGCTTTGAC TCCAGGATCC CTGGAGTGCC 180
 15 TTGGTGTTC AAGCCCCTGA CTAGGAAGAC AGAATGCACC TTCTGAGGCA CCTCCAGCTG 240
 CCCCTGGGAT GCAGGCTGAG CACCCTTGCC CGGCTGTGAT TGCTGCCAGG CACTGTTTCAT 300
 CTCAGTTTTT CTGTCCCTTT GCTCCCGGCA AGCTTTCTGC TGAAAGTTCA TATCTGGAGC 360
 CTGATGTCTT AACGAATAAA GGTCCCATGC TCCACCCGAA AAA 403

(2) INFORMATION FOR SEQ ID NO:4:

- 20 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 10 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

- 25 (ii) MOLECULE TYPE: genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

GGGCGGCCGC 10

(2) INFORMATION FOR SEQ ID NO:5:

- 30 (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 21 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: single
 (D) TOPOLOGY: linear

- (ii) MOLECULE TYPE: genomic DNA

- 35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

GTACATTCTG TCTCTTGCA A 21

(2) INFORMATION FOR SEQ ID NO:6:

- 47 -

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 24 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
5 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: genomic DNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:
TAACCCTGCT GCTGCTGGTC CTGG 24
- (2) INFORMATION FOR SEQ ID NO:7:
- 10 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- 15 (ii) MOLECULE TYPE: genomic DNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:
GTTTGCGTGC TGCCATGGAG A 21
- (2) INFORMATION FOR SEQ ID NO:8:
- 20 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: genomic DNA
- 25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
CCGCAATTAG AACAGCCTTG T 21
- (2) INFORMATION FOR SEQ ID NO:9:
- 30 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: genomic DNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
35 GCAGTGTAAC AACCGTGGTT GCTGC 25
- (2) INFORMATION FOR SEQ ID NO:10:
- 40 (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: genomic DNA

- 48 -

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

TGACCCCTGTG TCATCACCCCT GGC 23

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 20 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

CGGCTGCTCT GATGCCCGCC 20

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- 15 (A) LENGTH: 21 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

20 GCCGGCCACA GTCGATGAAT C 21

(2) INFORMATION FOR SEQ ID NO:13:

(i) SEQUENCE CHARACTERISTICS:

- 25 (A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

GAGAGGTTGC TGTTTTGATG ACA 23

30 (2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

- 35 (A) LENGTH: 23 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: genomic DNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

GCCAAGTCTT GATGTAGCCA GTT 23

(2) INFORMATION FOR SEQ ID NO:15:

- 49 -

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 60 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

Glu Ala Gln Thr Glu Thr Cys Thr Val Ala Pro Arg Glu Arg Gln Asn
 1 5 10 15
 Cys Gly Phe Pro Gly Val Thr Pro Ser Gln Cys Ala Asn Lys Gly Cys
 10 20 25 30
 Cys Phe Asp Asp Thr Val Arg Gly Val Pro Trp Cys Phe Tyr Pro Asn
 35 40 45
 Thr Ile Asp Val Pro Pro Glu Glu Glu Cys Glu Phe
 50 55 60

15 (2) INFORMATION FOR SEQ ID NO:16:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 62 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

Glu Lys Pro Ala Ala Cys Arg Cys Ser Arg Gln Asp Pro Lys Asn Arg
 1 5 10 15
 Val Asn Cys Gly Phe Pro Gly Ile Thr Ser Asp Gln Cys Phe Thr Ser
 20 25 30
 Gly Cys Cys Phe Asp Ser Gln Val Pro Gly Val Pro Trp Cys Phe Lys
 35 40 45
 Pro Leu Pro Ala Gln Glu Ser Glu Glu Cys Val Met Glu Val
 50 55 60

30 (2) INFORMATION FOR SEQ ID NO:17:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 59 amino acids
 (B) TYPE: amino acid
 (D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

Gln Glu Phe Val Gly Leu Ser Pro Ser Gln Cys Met Ala Pro Thr Asn
 1 5 10 15
 Val Arg Val Asp Cys Asn Tyr Pro Thr Val Thr Ser Glu Gln Cys Asn
 20 25 30
 40

- 50 -

Asn Arg Gly Cys Cys Phe Asp Ser Ser Ile Pro Asn Val Pro Trp Cys
 35 40 45

Phe Lys Pro Leu Gln Glu Thr Glu Cys Thr Phe
 50 55

5 (2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 74 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

10 (ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

Met Leu Gly Leu Val Leu Ala Leu Leu Ser Ser Ser Ser Ala Glu Glu
 1 5 10 15

15 Tyr Val Gly Leu Ser Ala Asn Gln Cys Ala Val Pro Ala Lys Asp Arg
 20 25 30

Val Asp Cys Gly Tyr Pro His Val Thr Pro Lys Glu Cys Asn Asn Arg
 35 40 45

Gly Cys Cys Phe Asp Ser Arg Ile Pro Gly Val Pro Trp Cys Phe Lys
 50 55 60

20 Pro Leu Thr Arg Lys Thr Glu Cys Thr Phe
 65 70

- 51 -

What is claimed is:

1. A polypeptide comprising a trefoil polypeptide, or a biologically active fragment thereof, for treating or inhibiting the formation of lesions.
- 5 2. The polypeptide of claim 1, wherein said trefoil polypeptide is a human trefoil polypeptide.
3. The polypeptide of claim 1, wherein said trefoil polypeptide is intestinal trefoil polypeptide (ITF).
4. The polypeptide of claim 1, wherein said trefoil
10 polypeptide is spasmodic polypeptide (SP).
5. The polypeptide of claim 1, wherein said trefoil polypeptide is PS2.
6. The polypeptide of claim 1, wherein said lesion is within the alimentary canal.
- 15 7. The polypeptide of claim 6, wherein said lesion is within the mouth of the patient.
8. The polypeptide of claim 6, wherein said lesion is in the esophagus of the patient.
9. The polypeptide of claim 6, wherein said lesion is in
20 the stomach of the patient.
10. The polypeptide of claim 6, wherein said lesion is in the intestine of the patient.
11. The polypeptide of claim 6, wherein said patient is receiving radiation therapy for the treatment of cancer.

- 52 -

12. The polypeptide of claim 6, wherein said patient is receiving chemotherapy for the treatment of cancer.
13. The polypeptide of claim 6, wherein said patient is receiving a drug that damages said alimentary canal.
- 5 14. The polypeptide of claim 6, wherein said patient is suffering from a digestive disorder.
15. The polypeptide of claim 14, wherein said digestive disorder is non-ulcer dyspepsia.
16. The polypeptide of claim 14, wherein said digestive
10 disorder is gastritis.
17. The polypeptide of claim 14, wherein said digestive disorder is gastro-esophageal reflux disease.
18. The polypeptide of claim 14, wherein said digestive disorder is a peptic ulcer or duodenal ulcer.
- 15 19. The polypeptide of claim 6, wherein said administration is oral administration.
20. The polypeptide of claim 6, wherein said oral administration comprises administration of about
10 milligrams to about 100 milligrams of said polypeptide.
- 20 21. The polypeptide of claim 1, wherein said lesion is within a tissue other than a tissue in the alimentary canal.
22. The polypeptide of claim 21, wherein said tissue is the skin.

- 53 -

23. The polypeptide of claim 22, wherein said administration is topical administration.

24. The polypeptide of claim 23, wherein said topical administration comprises administration of an ointment
5 containing about 1 mg/ml to about 10 mg/ml of the polypeptide.

25. The polypeptide of claim 21, wherein said tissue comprises the corneal surface of the eye.

26. The polypeptide of claim 21, wherein said tissue
10 comprises a tissue in the respiratory tract.

27. The polypeptide of claim 21, wherein said tissue comprises a tissue in the genitourinary tract.

28. A composition comprising a trefoil polypeptide for treating or inhibiting the formation of lesions in the
15 tissue of a patient.

29. The use of the composition of claim 28 in the manufacture of a medicament for the treatment of lesions in the tissue of a patient.

30. The trefoil polypeptide of claim 1, said polypeptide
20 further comprising a marker.

31. The trefoil polypeptide of claim 30, wherein said marker comprises an imaging agent.

32. A method for detecting an ITF receptor in a tissue, said method comprising contacting said tissue with
25 detectably labelled trefoil polypeptide and measuring the

- 54 -

level of detectably labelled trefoil polypeptide bound to said tissue.

33. A method for detecting a trefoil polypeptide in a tissue, said method comprising contacting said tissue with
5 an antibody that specifically binds said trefoil polypeptide.

1/9

17 gaagtttgcg tgctgcc
62 atg gag acc aga gcc ttc tgg ata acc ctg ctg gtc ctg gtt
107 gct ggg tcc tcc tgc aaa gcc cag gaa ttt gtt ggc cta tct cca
152 agc caa tgt atg atg gcg cca aca aat gtc agg gtg gac tgt aac tac
197 ccc act gtc aca tca gag cag tgt aac aac cgt ggt tgc tgt ttt
242 gac tcc agc atc cca aat gtg ccc tgg tgc ttc aaa cct ctg caa
260 gag aca gaa tgt aca ttt
310 tgaagctgtc caggctccag gaaggaggct ccacaccctg gactcttgct
360 gatggtagtg gccagggta acaactcacc ctgatctgct ccctcgcgcc
410 ggccaatata ggagctggga gtccagaaga ataaagacct tacagtcagc
431 acaaggctgt tctaattgcg g
(SEQ ID NO: 1)

FIG. 1

2/9

| | | | |
|---|----|----|----|
| Met Glu Thr Arg Ala Phe Trp Ile Thr Leu Leu Val Leu Val | 5 | 10 | 15 |
| Ala Gly Ser Ser Cys Lys Ala Gln Glu Phe Val Gly Leu Ser Pro | 20 | 25 | 30 |
| Ser Gln Cys Met Ala Pro Thr Asn Val Arg Val Asp Cys Asn Tyr | 35 | 40 | 45 |
| Pro Thr Val Thr Ser Glu Gln Cys Asn Asn Arg Gly Cys Cys Phe | 50 | 55 | 60 |
| Asp Ser Ser Ile Pro Asn Val Pro Trp Cys Phe Lys Pro Leu Gln | 65 | 70 | 75 |
| Glu Thr Glu Cys Thr Phe | | | |

(SEQ ID NO: 2) 80

FIG. 2

SUBSTITUTE SHEET (RULE 26)

3/9

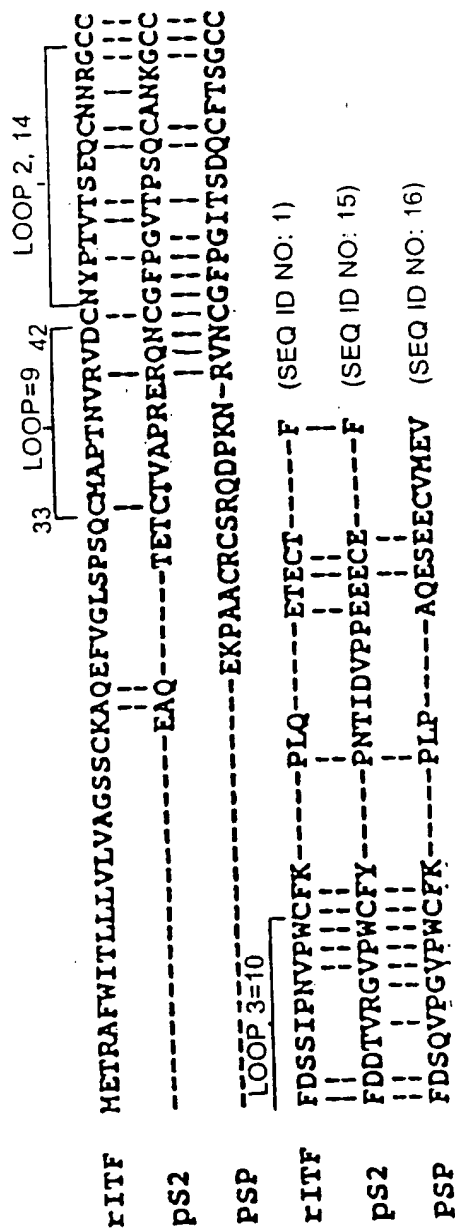


FIG. 3

4/9

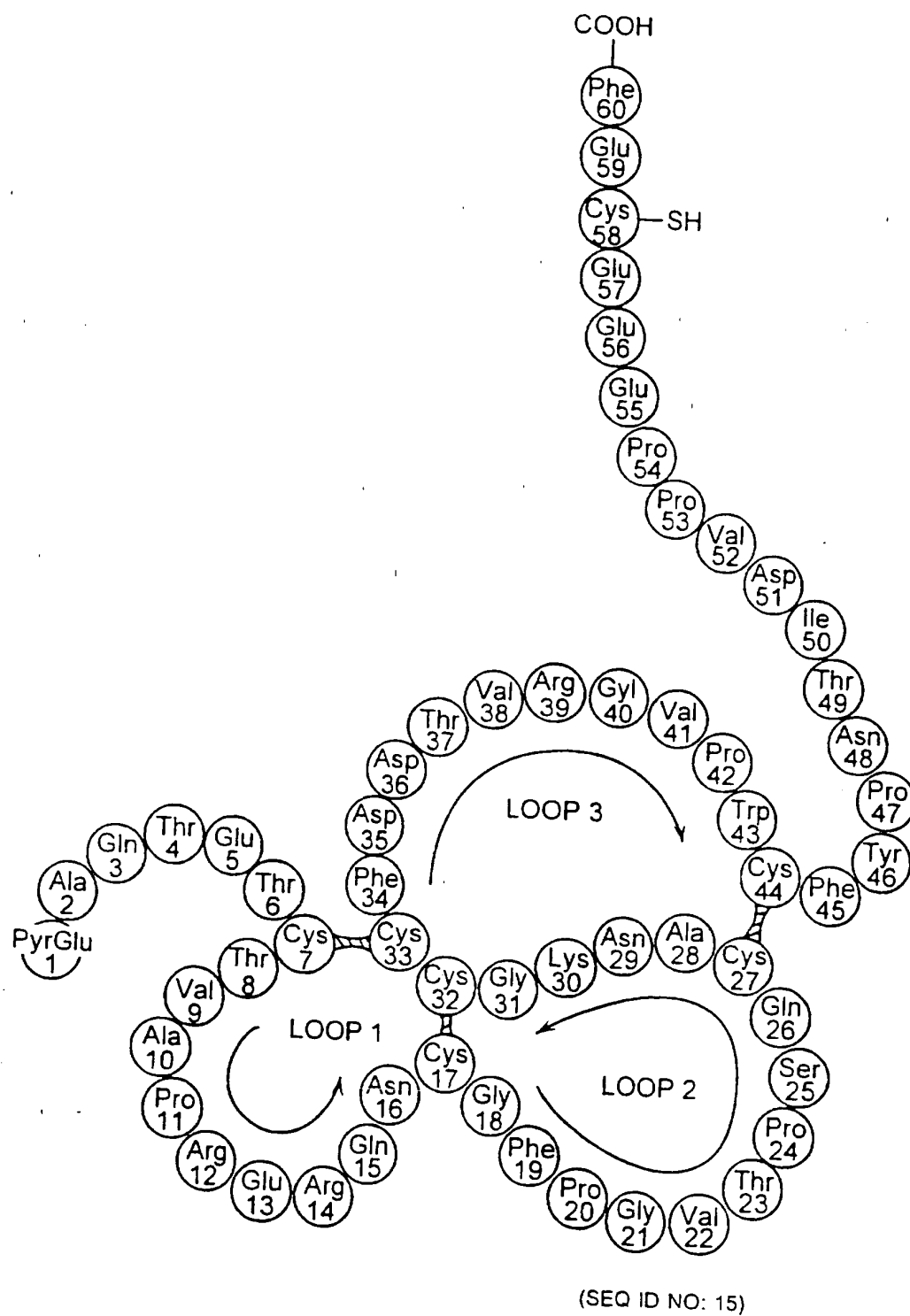
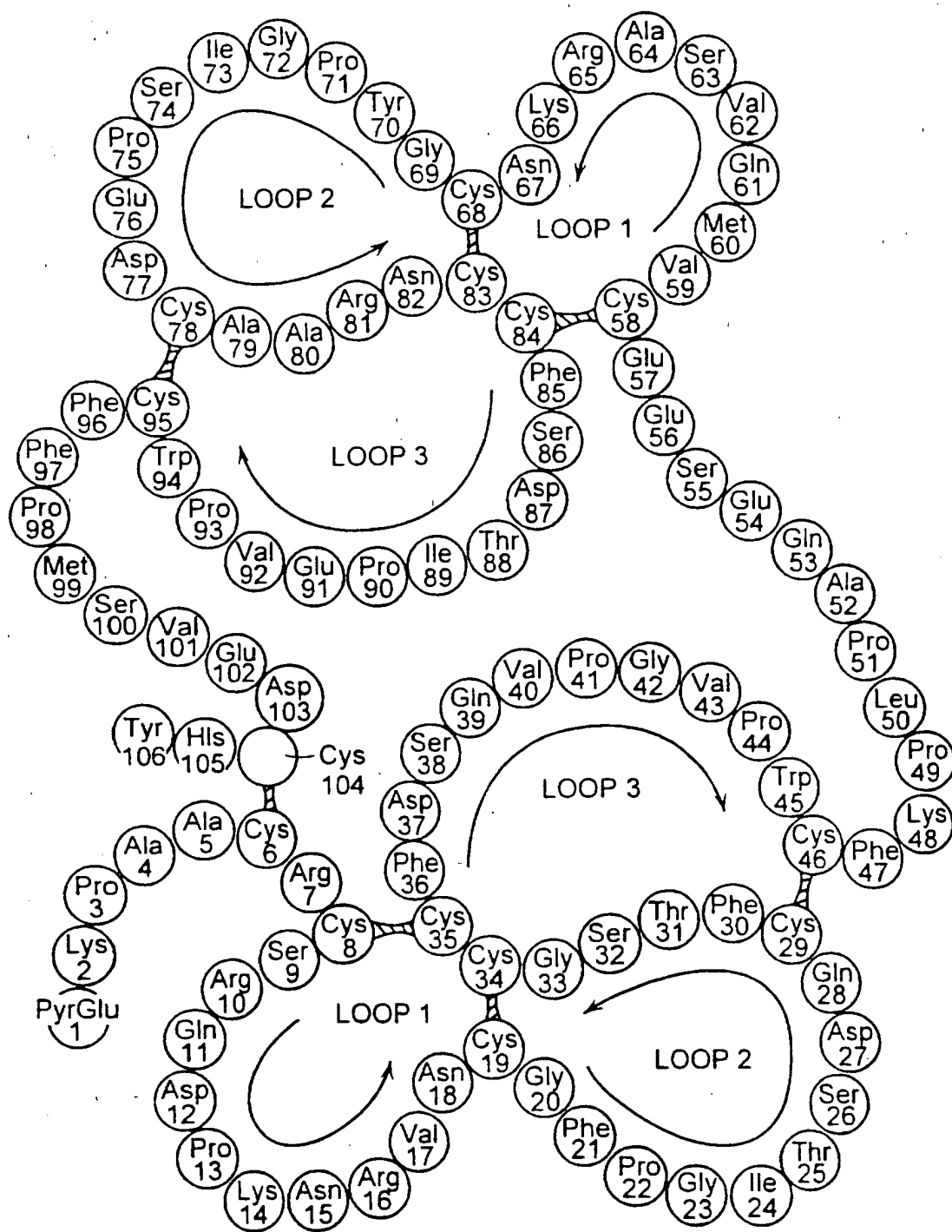


FIG. 4A

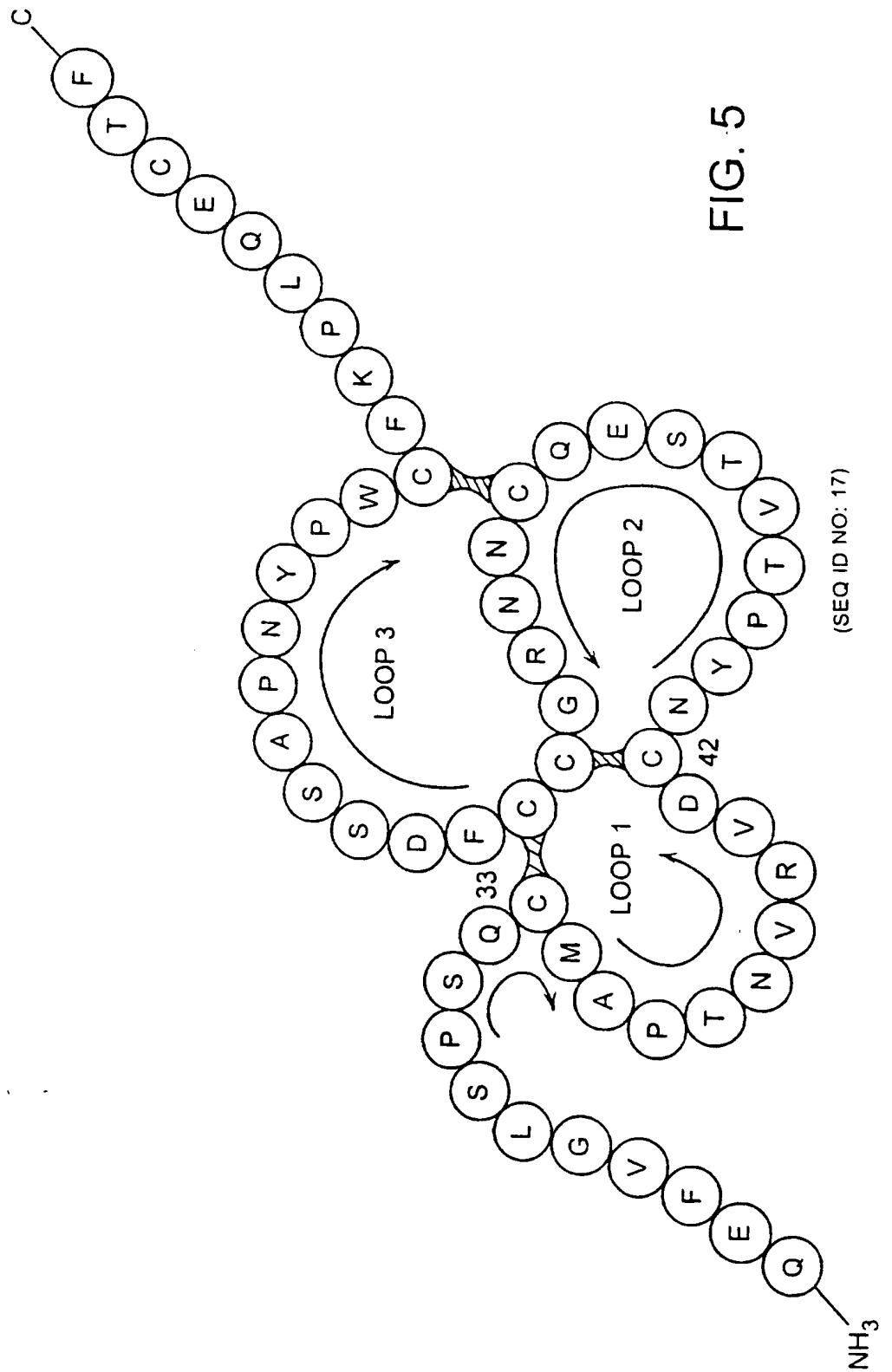
SUBSTITUTE SHEET (RULE 26)

5/9



(SEQ ID NO: 16)

FIG. 4B



7/9

```
gatgctgggctggtcctggccttgctgtcctccagctctgctgaggagtaagtggcct 60
M L G L V L A L L S S S A E E Y V G L
gtctgcaaaccaagtgtgccgtgccggccaaggacaggtggactgggctacccccatgt 120
S A N Q C A V P A K D R V D C G Y P H V
caccccaaggagtgcacaacccgggctgctgttgaactccaggatccctggagtgcc 180
T P K E C N N R G C C F D S R I P G V P
ttggtgttcaagcccctgactaggaagacagaatgcaaccttctgaggcacctccagctg 240
W C F K P L T R K T E C T F *
cccctgggatgcaggctgagcacccttgcccggctgtgattgctgccaggcactgttcat 300
ctcagtttttctgtcccttctgtcccggaagcttctgtgaaagtcatatctggagc 360
```

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ctgatgtcttaacgaataaagggtcccatgtctccaccgAAAAA  SEQ ID NO:3
-----+-----+-----+-----+-----+403  SEQ ID NO:18
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FIG. 6

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8/9

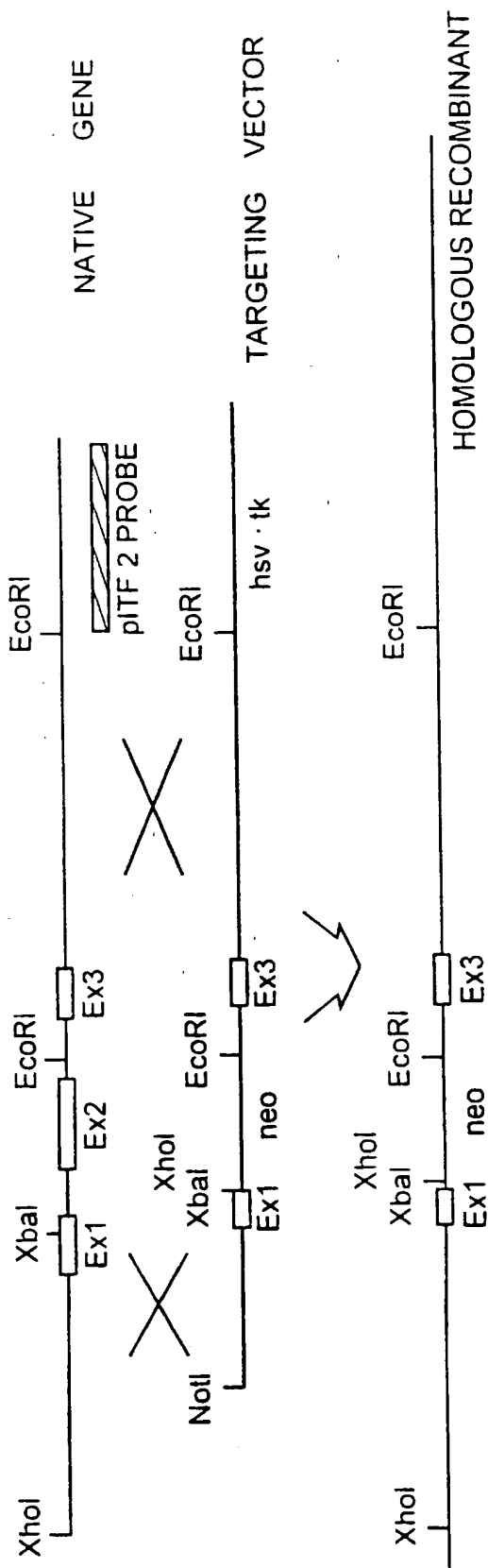


FIG. 7

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9/9

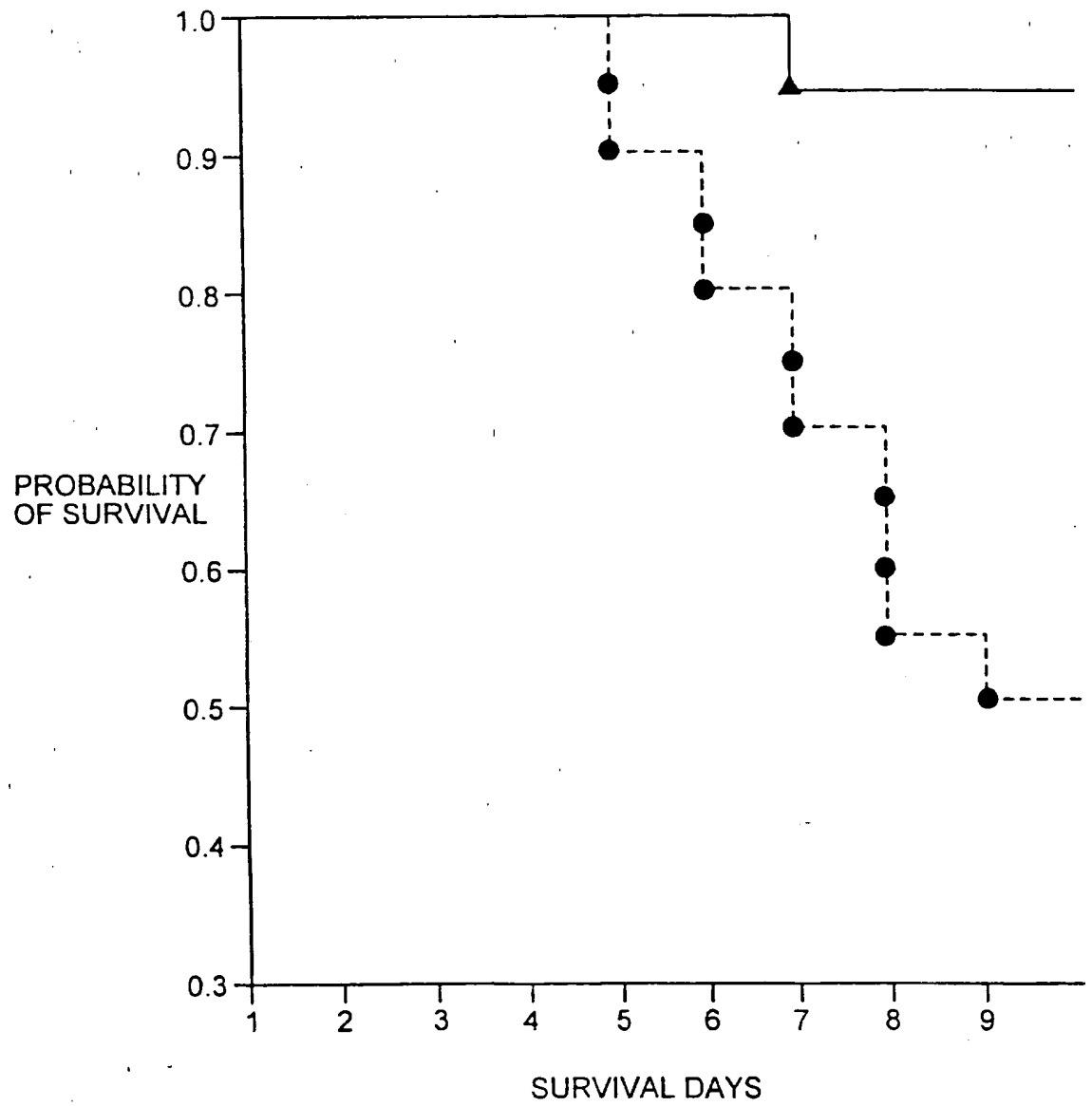


FIG. 8

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US97/06004

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) : A61K 39/00; C07K 14/47; G01N 33/567

US CL : 424/184.1; 435/7.2; 530/350

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/184.1; 435/7.2; 530/350

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, CA, MEDLINE, BIOSIS, EMBASE

search terms: intestinal trefoil polypeptide

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category* | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|--------------|---|------------------------|
| X | PODOLSKY et al. Identification of Human Intestinal Trefoil Factor. The Journal of Biological Chemistry. 25 March 1993, Vol. 268, No. 9, pages 6694-6702, see entire document. | 1-27 |
| X -- Y | BABYATSKY, M.W. et al. Trefoil Peptides Protect Against Ethanol and Indomethacin Induced Gastric Injury in Rats. Gastroenterology. April 1994, Vol. 106, No. 4, page A43, first column, second abstract, see entire abstract. | 1-29 ----- 30-33 |



Further documents are listed in the continuation of Box C.



See patent family annex.

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| *P* document published prior to the international filing date but later than the priority date claimed | |

Date of the actual completion of the international search

09 JUNE 1997

Date of mailing of the international search report

24 JUL 1997

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